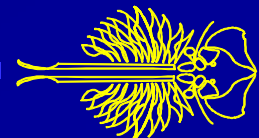


# Genetic implications in the production of rotifers in commercial finfish hatcheries

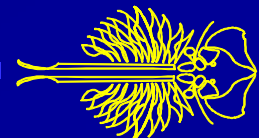
EU project

October 2002-2005

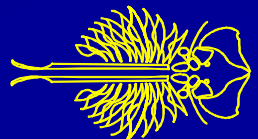
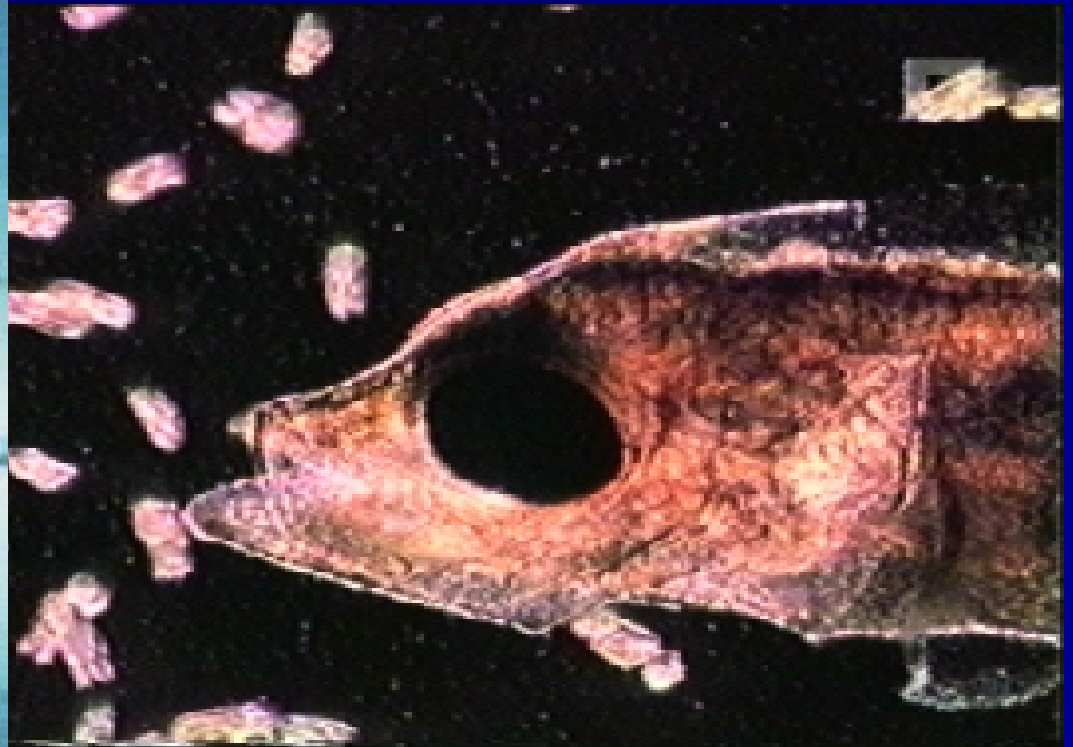
S. Dooms<sup>1</sup>, K. R. Dierckens<sup>2</sup>, A. Abatzopoulos<sup>3</sup>, A. Triantaphyllidis<sup>3</sup>,  
Y. Olsen<sup>4</sup>, T. De Wolf<sup>5</sup>, D. Delbare<sup>1</sup>, P. Bossier<sup>2</sup> & P. Sorgeloos<sup>2</sup>



- 1: Ministry of Agriculture, Centre of Agriculture Research, Sea Fisheries Department, Belgium
- 2: Universiteit Gent, Laboratory of Aquaculture & Artemia Reference Center, Belgium
- 3: Aristotle University of Thessaloniki, Department of Genetics, Development and Molecular Biology, Greece
- 4: Norwegian University of Science and Technology, Norway
- 5: Maricoltura di Rosignano Solvay srl, Italy

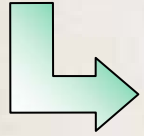


# Fish and Crustacean culture depend on the production of rotifers



## Rotifer cultures:

### Conditions in favour of parthenogenetic cycle



**Faster reproduction**

**Ability of bioencapsulation**

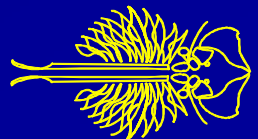
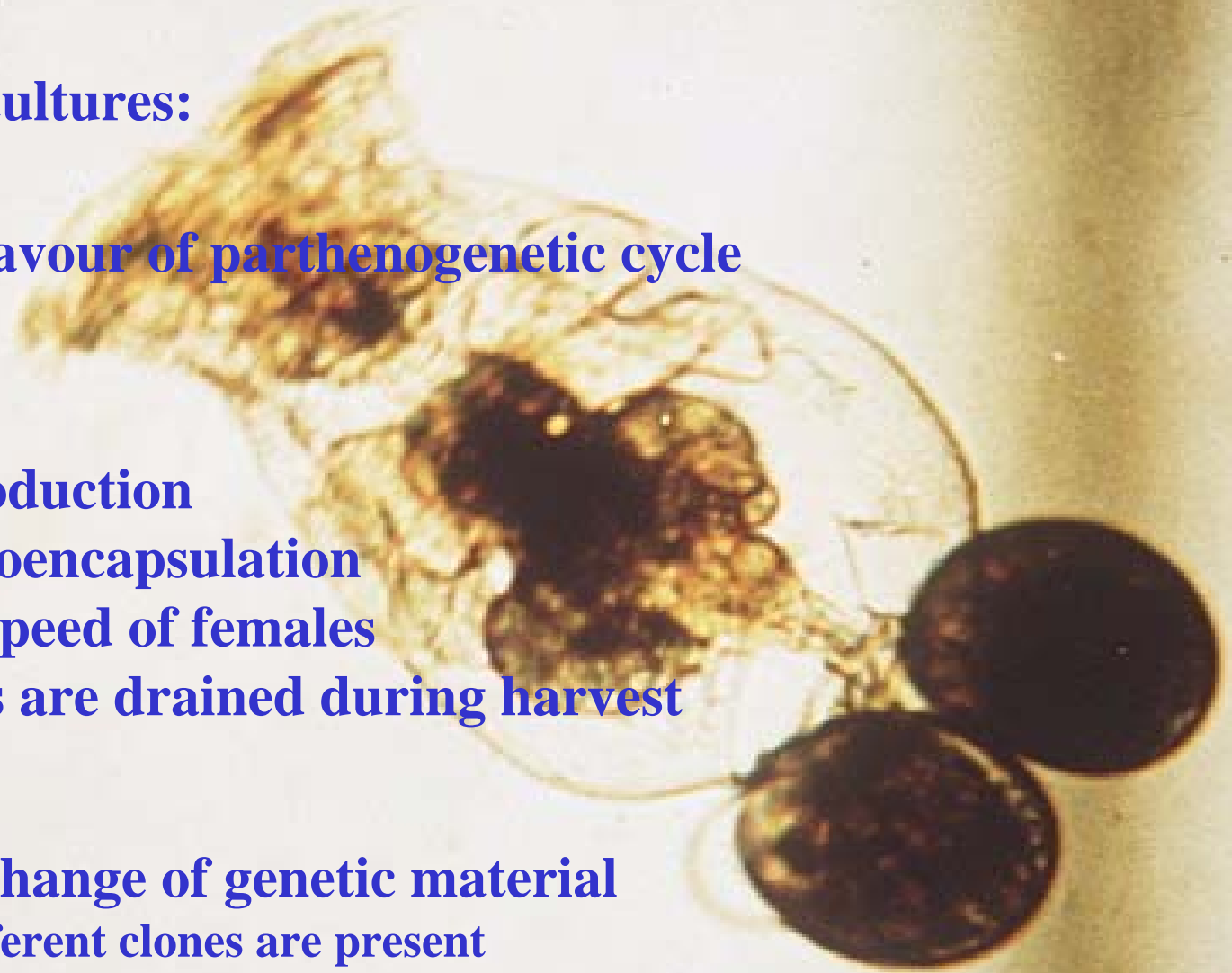
**Swimming speed of females**

**Resting eggs are drained during harvest**



**Little exchange of genetic material**

**Even if different clones are present**



**PROBLEM: Unpredictable culture crashes**



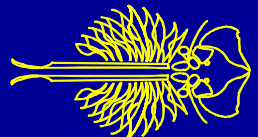
**Unreliable live feed source**



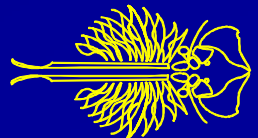
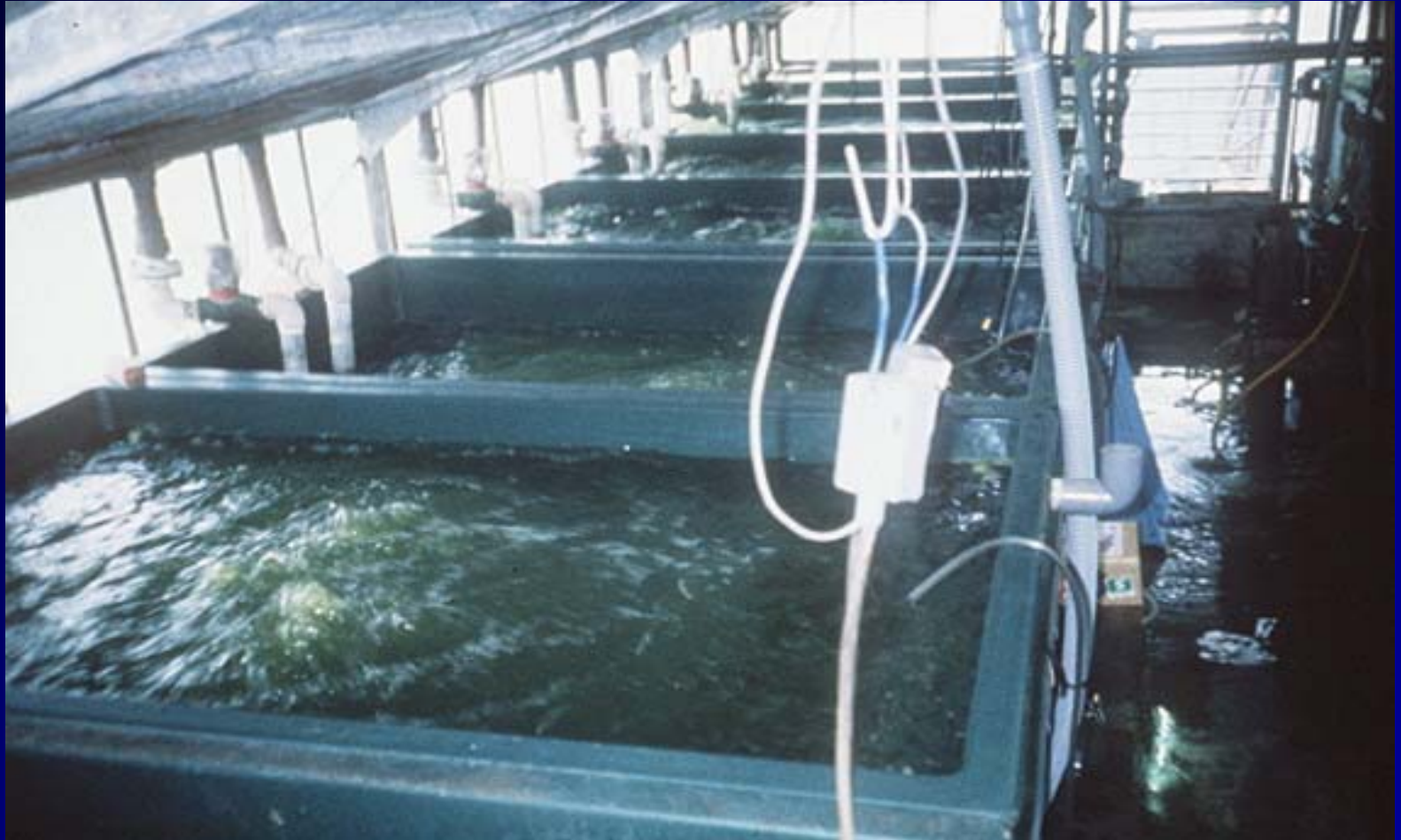
**Unpredictable fish/shrimp larviculture**



**Unpredictable income**



**Solution:** **Farmers double the amount of rotifer culture tanks**  
**Nearby hatcheries help each other in case of emergency**

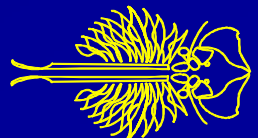


**Solutions: only dealing with consequences not with causes**

**ROTIGEN project**

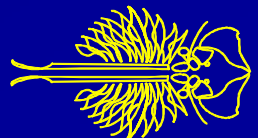


**Shed light on the possible causes of rotifer crashes**



## Hypotheses:

- 1) **Does the controlled mass culture of rotifers lead to an impoverishment in the genetic diversity of the cultured rotifers?**
- 2) **Does this fact in its turn make the rotifer culture more susceptible to crashes if there is a change, controlled or uncontrolled, in the biotic or abiotic conditions prevailing in the culture?**



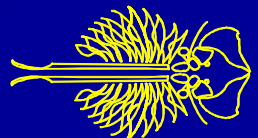
# Overview (Part 1)

**Topic 1: Genetic characterisation of rotifer clones**

**Topic 2: Effect of culture methods on the genetic stability of rotifers**

**Topic 3: Effects of environmental conditions on the genotypic diversity of cultured rotifers**

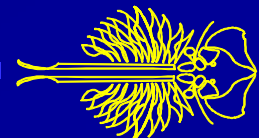
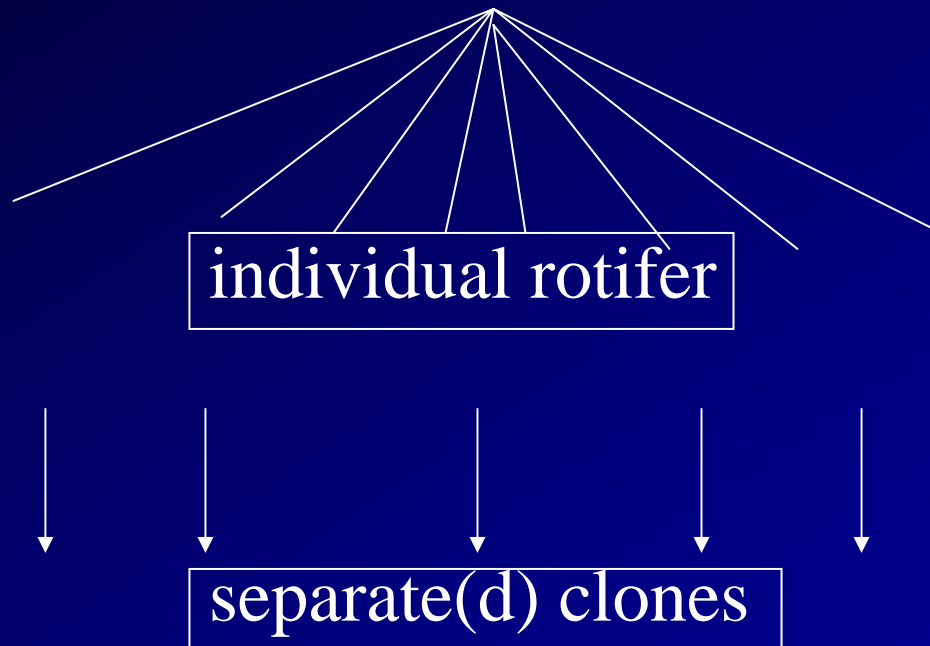
**Topic 4: Effect of microbial stability on the genotypic diversity of cultured rotifers**



# Topic 1: Genetic characterisation of rotifer clones

## Isolation and maintenance of rotifer clones

Samples from different hatcheries/labs



## Maintenance of living rotifer clones:

Falcon tubes of 50 ml

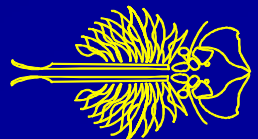
Culture period: 1 week

Autoclaved sea water

Feed: *Chlorella sp.*

Salinity: 25 ppt

Temp.: 25°C



Standardisation of protocols for: sampling & storage  
transport

Building of a reference collection

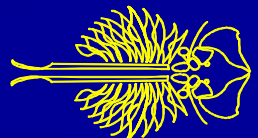
Clones genetically characterised with:

16 S rDNA gene

HSP 60 gene

Microsatellite markers

(Results on the genetic characterisation: see poster section)



## Topic 2: Effect of culture methods on the genetic stability of rotifers

### 1) Interclonal selection for different culture methods

#### Batch culture

(Belgium): 80 L

4 day culture period

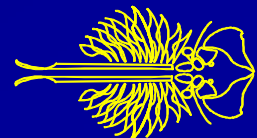
Constant volume

25 ppt, 25°C

Feed: different kinds

Inoculation: 500 ind/ml

or 50 ind/ml



Recirculation culture (Belgium) 80 L

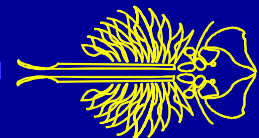
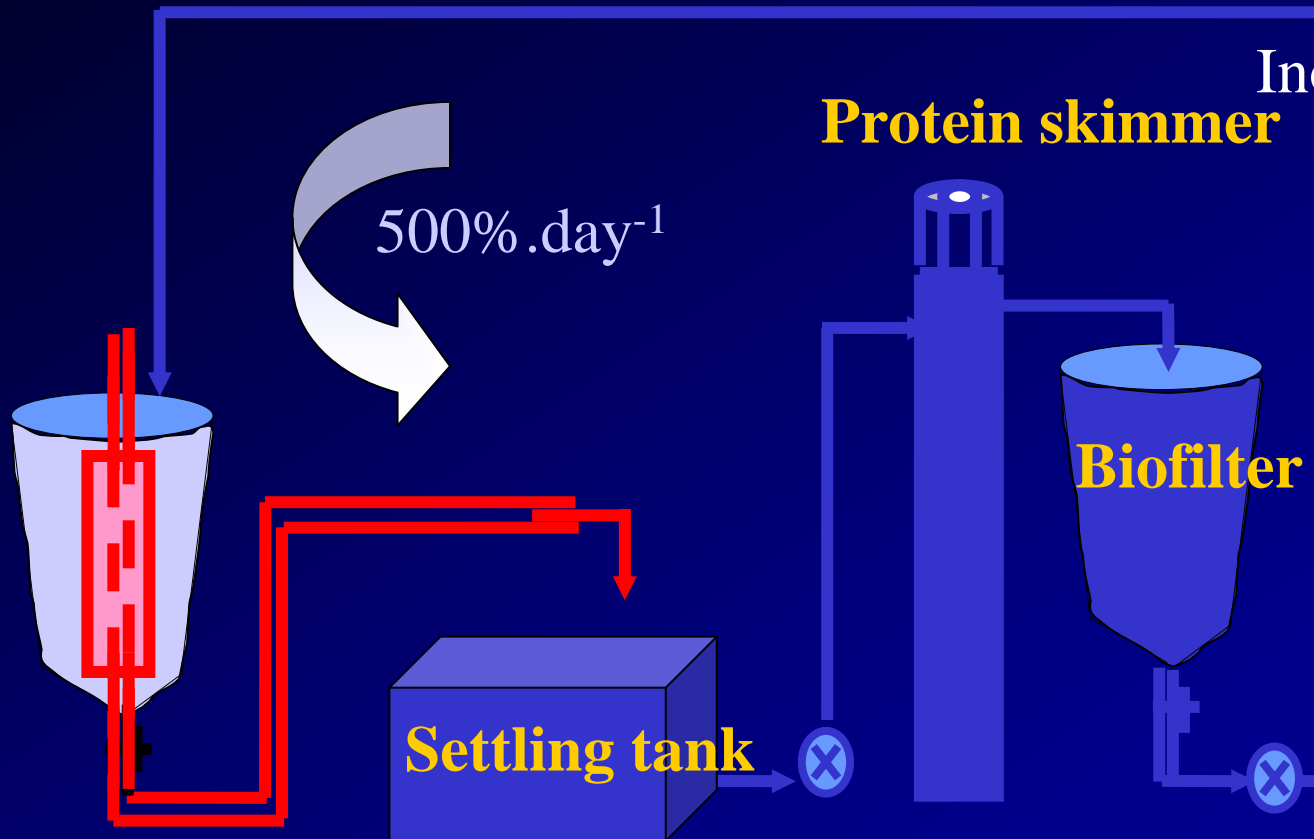
3 – 4 weeks culture period

25 ppt, 25°C

feed: different diets

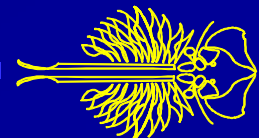
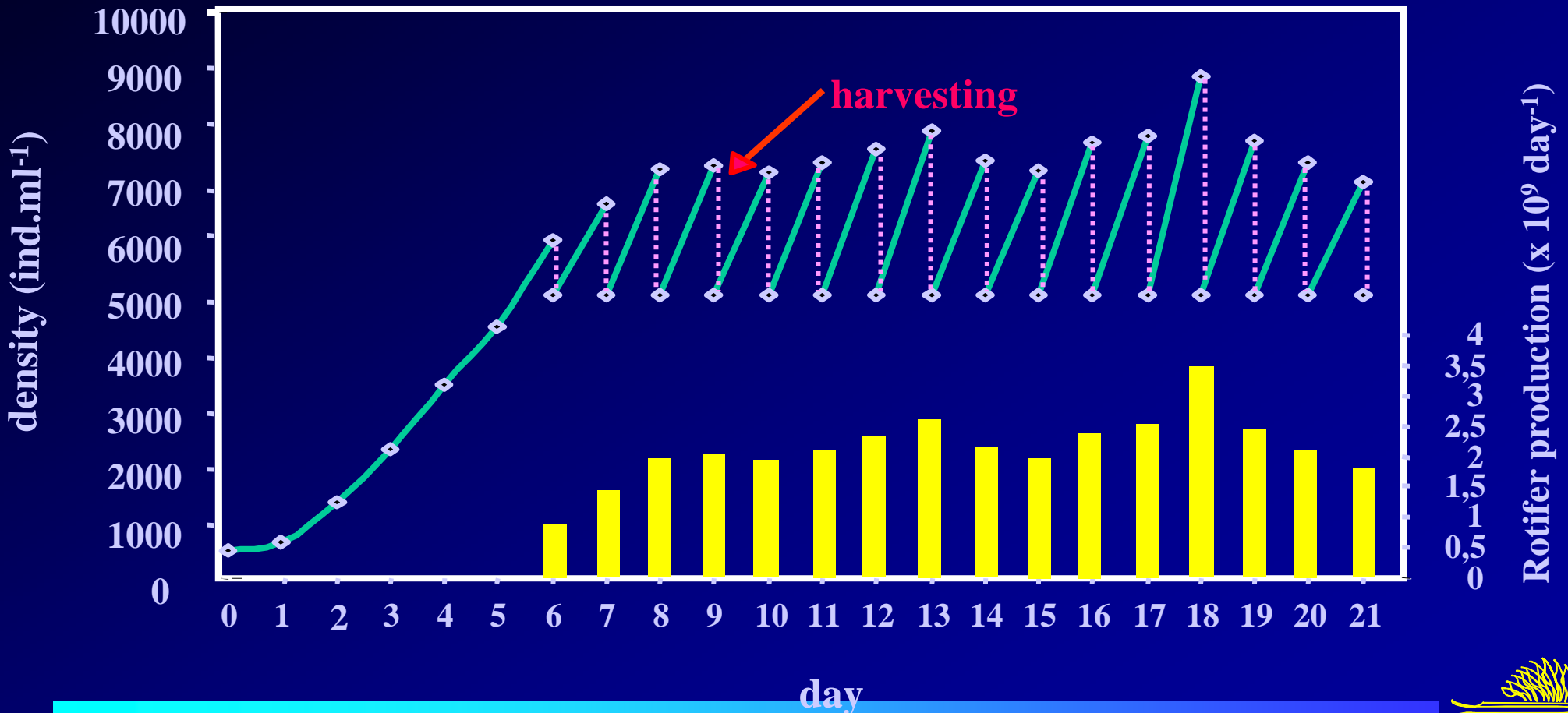
Inoculation: 500 rotifers/ml

or 50 rotifers/ml



# Semi-continuous culture: Norway

Constant rotifer density: daily harvests  
+ addition of new water



A mixture of identifiable rotifer clones at equal inoculation densities



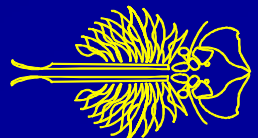
Different culture methods



**Sampling:** Start, after every run (for batch), every week (for recirc)



Selection of rotifer clones dependent on the culture method



# Topic 3: Effects of environmental conditions on the genotypic diversity of cultured rotifers

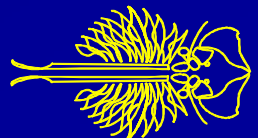
Axenic parthenogenetic eggs → Axenic cultures

Stable conditions

temperature

feed

salinity



# Topic 3: Effects of environmental conditions on the genotypic diversity of cultured rotifers

Axenic parthenogenetic eggs → Axenic cultures

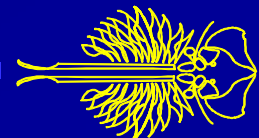
Variable conditions

Temperature

feed

Within 1 culture cycle

Variability between culture cycles



## Topic 4: Effect of microbial stability on the genotypic diversity of cultured rotifers

### 1) Effect of selected bacterial strains on the interclonal selection

Axenic rotifer culture + 5 bacteria

Feed: Gamma-irradiated

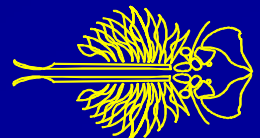
axenic culture of algae

Sampling: daily: rotifer density + egg ratio

weekly: genetical analyses

└ community:PCR-DGGE

ARC + NTNU: different salinity + temperature



## 2) Effect of selected bacterial strains on the interclonal selection in xenic cultures

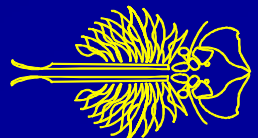
Back to reality:

xenic rotifer culture + 5 bacterial strains

**ARC:** batch cultures

**NTNU:** semi-continuous culture

**Expected results:** Protocol for addition of selected bacteria resulting in high production and low selection



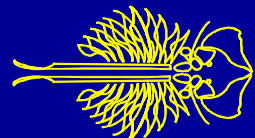
## Overview (Part 2)

**Topic 5: Verification of genetic selection under pilot scale conditions**


**Topic 6: Sexual reproduction to provide resting eggs of selected strains**

**Topic 7: Follow up of cultures in commercial hatcheries**

**Final : Workshop for the industry**



## Topic 5: Verification of genetic selection under pilot scale conditions

Strains from Mediterranean hatcheries  ARC  
NTNU

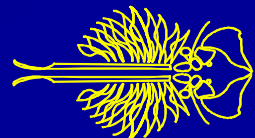
ARC: batch or recirculation culture

NTNU: semi-continuous culture

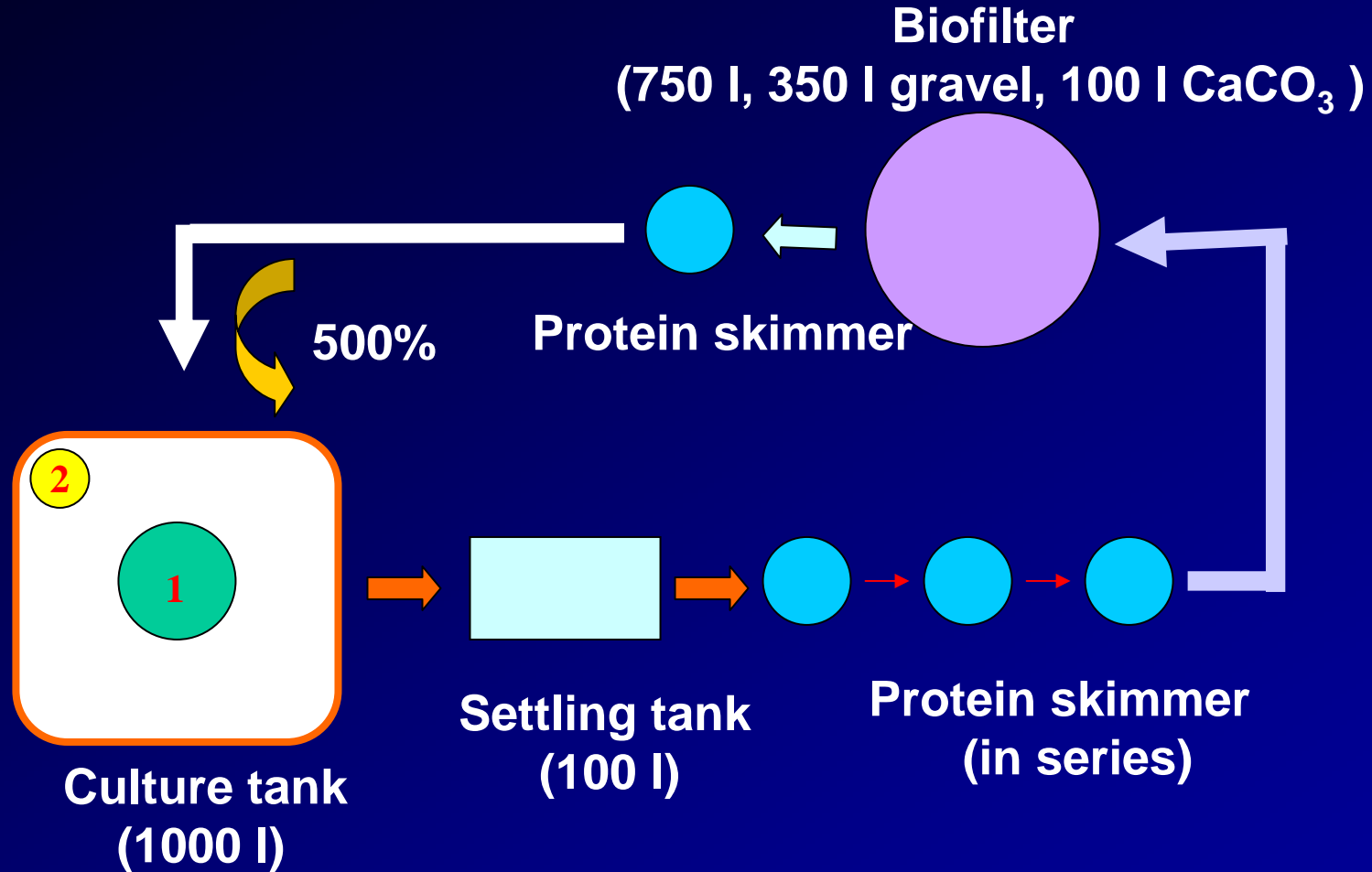
Methods: according to WP2

**Deliverables:** Adaptability of rotifers from commercial hatcheries to other culture conditions

**Expected results:** Data on clonal selection in rotifer populations in commercial cultures

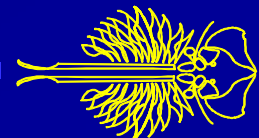


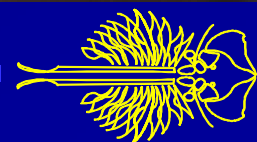
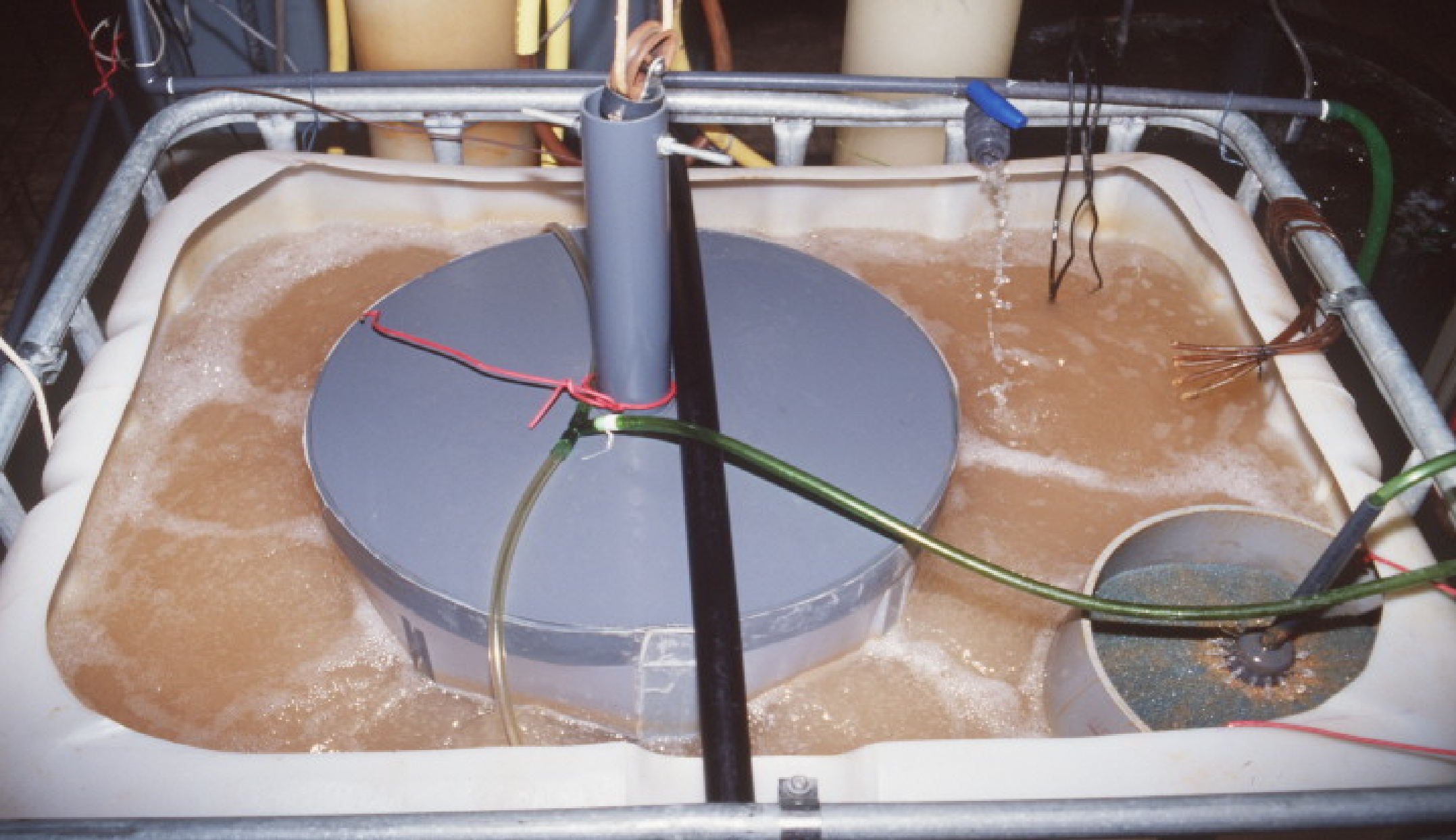
# Schematic outline of the pilot scale system



1 = filter (30 µm)

2 = air water lift





## **Topic 6: Sexual reproduction to provide resting eggs of selected strains**

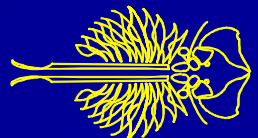
### **1) Methodology for resting egg production, harvest and storage**

Temperature, salinity, feed quantity, photoperiod, population density

Triggers will vary among the different clones

### **2) Selection of competent strains**

Screening of genetically characterized clones for ability of resting egg production



### 3) Genetic characterisation of resting egg banks

Resting eggs produced in 2) will be checked for diversity on  
microsatelite scale

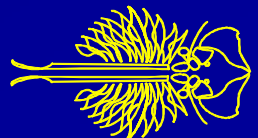
### 4) Application of resting eggs for aquaculture

Resting eggs as inoculum  
as remedy against genetic impoverishment

Evaluation of the use of resting eggs for aquaculture

### 5) Cryopreservation of amictic eggs

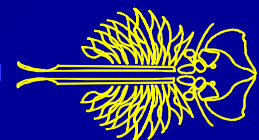
Advantage: easier production  
genetically identical



## Topic 7: Follow up of cultures in commercial hatcheries

Rotifer samples from MRS    from stockcultures  
from start of cultures  
at onset of a crash or at the end of a  
culture cycle

Rotifer samples from other hatcheries through connections of the RTD  
partners (Greece, France, Norway)



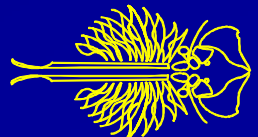
## Final : Workshop for the industry



ARC will organise a workshop on mass production of rotifers and the genetic consequences

AUTH and NTNU will also invite farmers

**Expected results:** Dissemination of the results to the industry



A close-up photograph showing a human hand reaching over a tray filled with numerous small, translucent, teardrop-shaped organisms, which are Artemia nauplii. The organisms are densely packed and appear to be in various stages of development. The background is a dark, slightly textured surface, possibly the tray itself. The lighting is bright, highlighting the delicate structure of the organisms and the skin of the hand.

# The end

